

Procedure for Protein/Peptide Neutralization with EndoPrep™

1. Prepare sample by diluting to a concentration of 0.1 to 1.0 mg/ml with BDT™ Digestion Buffer.
 - BDT™ Digestion Buffer should account for at least 90% of the sample volume for proper protease activity.
 - If dilution is not possible, use diluted hydrochloric acid to lower the pH of the sample below 4.5.
 2. Remove an aliquot of 270 µl and transfer to a sterile, endotoxin-free borosilicate glass tube.
 3. Add 30 µl of BDT™ Protease Solution and mix by vortexing for 10 seconds.
 - A control sample containing 30 µl of BDT™ Digestion Buffer instead of BDT™ Protease Solution should be included with each set of reactions to determine baseline endotoxin content without digestion.
 4. Cover tube with Parafilm®.
 5. Incubate tube in a 37°C water bath.
 - Most proteins and peptides show maximum endotoxin activity after 60 minutes of treatment. Some samples may require longer incubation.
 - For examples of digestion times, refer to the BioDrech, Inc. EndoPrep™ Application Notes.
 - To verify complete sample digestion, polyacrylamide gel electrophoresis should be performed on digested samples.
 6. After digestion, dilute samples 1:100 in endotoxin-free water.
 - Dilution of only 1:10 is possible but should include proper validation that enzyme or digestion products do not interfere with the endotoxin detection assay at this concentration.
 7. Test with LAL or recombinant Factor C assay.
 - Samples treated with EndoPrep™ should be tested both with and without a positive product control (PPC).
 - Samples of BDT™ Digestion Buffer and BDT™ Protease Solution at equivalent concentrations should be tested alongside all samples as control.
- ### Procedure for EndoPrep™ Preparation and Storage
1. Upon receipt, store EndoPrep™ kit at 4°C.
 - Before preparation of BDT™ Protease Solution, the EndoPrep™ kit is stable for 2 years when properly stored.
 2. Before use, add 1 ml BDT™ Digestion Buffer to BDT™ Protease Solution bottle.
 3. Mix sample vigorously with vortexing for 5 minutes.
 4. Assure full solubilization by visual inspection.
 5. BDT™ Protease Solution should be stored at 4°C.
 - After preparation of BDT™ Protease Solution, the EndoPrep™ kit is stable for 3 months when properly stored.

Predictive Oncology was organized to develop and market products for detection, removal and neutralization of bacterial toxins.

Endotoxin Detection Products:
EndoPrep™ 20 reactions EDP-4001.01

Endotoxin Removal Products:
EndoBind-R™ 1 ml column EBR-3001.01
EndoBind-R™ 5 ml column EBR-3005.01
EndoBind-R™ Bulk resin Inquire

• For Research Use Only •

200 Riverchills Business Park
Suite 250
Birmingham, AL 35242
E-mail: orders@predictive-oncology.com

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Oncology®**

www.predictive-oncology.com

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- Easy to use in most samples.
- Requires 60 minutes for most samples.
- Works with LAL and recombinant Factor C assays.
- Removes inhibitory effect of endotoxins.
- Increases detection accuracy.

Advantages

U.S. Patent Pending

EndoPrep™ is a sample treatment system to remove the inhibitory effects of proteins and peptides and allow accurate detection and quantitation. The system consists of BDT™ Digestion Buffer and BDT™ Protease Solution. Following the provided protocol, both the LAL and recombinant Factor C endotoxin detection assays are compatible with the system components. The system components are compatible with both classical LAL and recombinant Factor C endotoxin detection assays.

Product Description



EndoPrep™

Protein/Peptide
Sample Treatment

Catalog No: EDP-4001

www.predictive-oncology.com

EndoPrep™ and Endotoxin

The majority of bacteria consists of lipopolysaccharide (LPS) on the outer membrane of gram-negative bacteria. LPS is also called endotoxin. (LPS) endotoxin also contains a "flag" or "tag" molecule. Sub-nanogram levels of endotoxin can alter the phagocytosis and immune response of cells including neutrophils, dendritic cells, hepatocytes, and macrophages. Because of this, the presence of endotoxin in a sample can affect the results of endotoxin detection. Endotoxin is a protein that is secreted by bacteria and is a major component of the cell wall. It is a highly active and stable molecule. Endotoxin is a protein that is secreted by bacteria and is a major component of the cell wall. It is a highly active and stable molecule. Endotoxin is a protein that is secreted by bacteria and is a major component of the cell wall. It is a highly active and stable molecule.

Endotoxin Detection in an Inhibitory Peptide Sample Using EndoPrep™

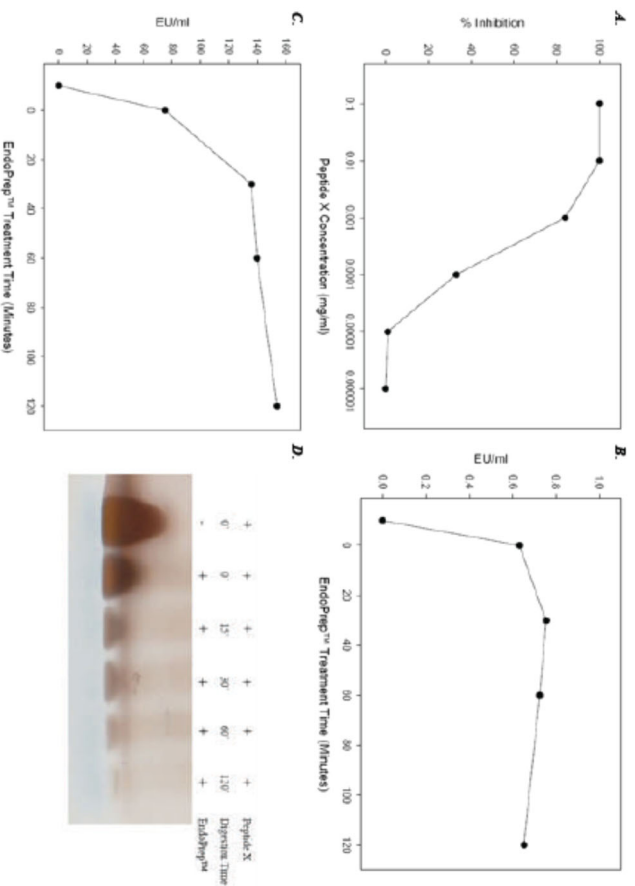


Figure 1. Treatment of Peptide X with EndoPrep™. (A) The PPC inhibition Assay shows extent of peptide dilution required to overcome inhibitory effect. (B) Recovery of a 1 EU/ml PPC in Samples of Peptide X before and after treatment with EndoPrep™. (C) Recovery of defined endotoxin contamination in a sample of Peptide X. (D) PAGE data showing Peptide X degradation with EndoPrep™ treatment.

Peptide X (proprietary) is a mixture of short cationic peptides shown to be extremely effective in a therapeutic setting. However, the cationic nature of Peptide X causes it to bind endotoxin and mask its activity from LAL and recombinant Factor C assays. The extent of this inhibition is shown in Figure 1A which details the requirement of a 2,000,000-fold dilution (to 10 ng/ml) for full endotoxin spike recovery. A sample of 0.1 mg/ml Peptide X was prepared in BDT™ Digestion Buffer containing 250 EU/ml exogenous endotoxin contamination. This sample was then incubated for various times with the BDT™ Protease Solution and endotoxin content was measured with the Lonza PyroGene® assay. Figure 1B shows the results of 1 EU/ml PPC recovery experiments for all digestion samples. Without digestion, no endotoxin was detected. After EndoPrep™ treatment, nearly 80% of the PPC was recovered, exceeding the standard 50-200% recovery requirements. Recovery of exogenous endotoxin contamination showed similar results (Figure 1C). Without treatment, none of the 250 EU/ml were detected. With treatment, over 150 EU/ml were detected. Figure 1D shows that Peptide X degradation from EndoPrep™ treatment corresponds to the removal of endotoxin masking.

For a more detailed explanation of EndoPrep™ protocol refer to the **BioDtech, Inc. EndoPrep™** Application Notes. This document outlines the use of EndoPrep™ in treating peptides and proteins, including bovine serum albumin, immunoglobulin and hemoglobin.

Endotoxin Detection in Hemoglobin Solution using EndoPrep™

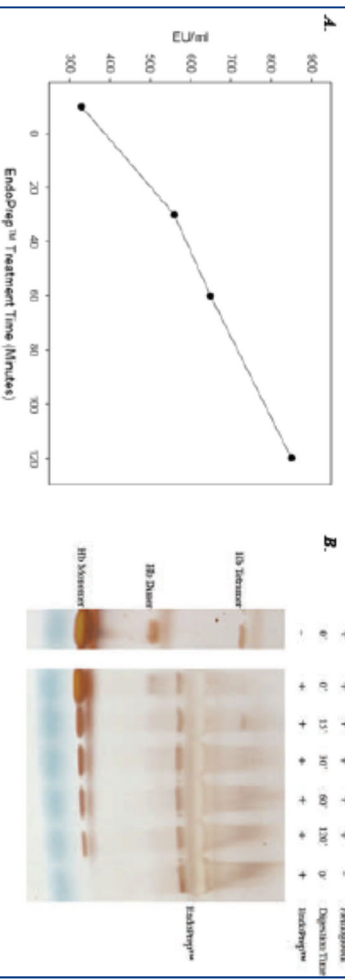


Figure 2. Treatment of Hemoglobin with EndoPrep™. (A) Recovery of endogenous endotoxin contamination in a sample of hemoglobin from bovine erythrocytes. (B) PAGE data showing hemoglobin degradation with EndoPrep™ treatment. The gel was run under reducing conditions resulting in three distinct hemoglobin populations. Locations of each hemoglobin population and the EndoPrep™ protease are indicated.

The example with Peptide X shows the use of EndoPrep™ in a sample that completely masks endotoxin activity. However, the EndoPrep™ system can also be used to increase the accuracy of endotoxin detection in protein samples that do not exhibit such dramatic effects. Hemoglobin is an example of a protein that is known to bind endotoxin and mask its activity. To increase the accuracy of endotoxin quantitation in a sample of hemoglobin, a 1 mg/ml sample was prepared in BDT™ Digestion Buffer. Previous experiments with this untreated hemoglobin stock showed that it contained about 300 EU/mg of endogenous endotoxin contamination. BDT™ Protease Solution was added to the hemoglobin sample and incubated at indicated times. Figure 2A shows that untreated hemoglobin measured at slightly more than 300 EU/mg, as previously reported. However, with increasing lengths of EndoPrep™ treatment, the amount of detectable endotoxin increased to about 650 EU/mg after one hour of treatment and 850 EU/mg after two hours of treatment. This indicates a 150% increase in detection and could be significant for biological samples. Figure 2B shows that hemoglobin digestion correlates to endotoxin liberation. The PAGE experiments were performed in reducing conditions resulting in monomer, dimer, and tetramer hemoglobin populations. With EndoPrep™ treatment, the dimer and tetramer populations are quickly degraded followed by continuous degradation of the monomer population over the entire two hour time course.

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